



FOUNDED  
1978

southern lepidopterists' news




VOL. 7 NO. 1  
MAY 1985

THE OFFICIAL PUBLICATION OF THE SOUTHERN LEPIDOPTERISTS' SOCIETY, ORGANIZED TO PROMOTE SCIENTIFIC INTEREST AND KNOWLEDGE RELATED TO UNDERSTANDING THE LEPIDOPTERA FAUNA OF THE SOUTHERN REGION OF THE UNITED STATES.

ANOTHER RECORD FOR AZALEA AS A FOODPLANT OF SATYRIUM LIPAROPS LIPAROPS (B & L)

By Bob Cavanaugh

ON March 16, 1985 while searching over the leaves of a wild azalea bush (Rhododendron canescens (Michaux)) I found a pale green, slug shaped larva which reared out to be a male Satyrium liparops liparops. The description of the larva contained in Klots' A FIELD GUIDE TO THE BUTTERFLIES is accurate for this subspecies. The larva turned a pretty lavender color three days prior to pupation. Pupation occurred on April 1st and the adult emerged on the morning of April 18th.

The azalea plant was in full bloom and the leaves were young and soft. The larva was positioned on top and centered on a rosette of leaves about 18 inches from the ground. The surrounding trees were Bitternut Hickory (Carya cordiformis (Wang)) which had just broken bud and beginning to push new growth. Water Oak is common in the vicinity but not close enough that the larva could have fallen or been blown onto the azalea bush.

[Ed. note] Azalea is a known foodplant of S. strigosum (Harris) according to Harris in BUTTERFLIES OF GEORGIA page 199. The closely related S. Kingi (Klots & Clench) has also been found in the wild feeding on azalea in the larval state. More attention should be given to "beating" wild azalea bushes to see how frequently this host is utilized by these two species.

1985 ANNUAL MEETING  
NEAR ST. MARKS NWR, FLORIDA

By Dave Baggett

I AM working towards setting up the 1985 annual meeting in or near St. Marks NWR (Inner Big Bend area along the NW Gulf Coast, about 40 miles south of Tallahassee). There has been very limited exploration in the area, and I think it is one that holds a great deal of promise. The date will be Labor Day weekend (Aug. 30 - Sept. 1). This could be a bit early for the area, but then again, it might just be right on target. The immediate area adjacent holds a lot of promise for several field trips, especially in conjunction with the long holiday weekend.

I will check things out during late May and will have full details worked out by mid-June. Let's begin to make plans now to get together Labor Day Weekend in unexplored areas of the Florida panhandle. We will have full details in the next newsletter including specific site, places to stay, and potential areas to explore. I am planning field demonstrations of various types of traps to serve as a practical workshop where members can learn how to use and assemble different types of traps. I will also see what I can do in terms of coming up with some slide shows from members. And, of course, we will feature our traditional meal together either at one of Apalachicola's seafood spots or a outdoor cookout, which ever proves most feasible.

## BOOK REVIEW

A FIELD GUIDE TO THE MOTHS OF EASTERN NORTH AMERICA, BY CHARLES V. COVELL, JR., HOUGHTON MIFFLIN Co., 1984, 496 pp., 62 PLATES (32 IN COLOR), \$18.95 / 13.95, HARD OR SOFT COVER.

HERE IS the text that many of us have been long waiting for, a companion guide for the popular Peterson Field Guide series to compliment A.B. Klots' Field Guide to the Butterflies East of the Great Plains (a revised edition forthcoming!). While many of us have worn the pages out of several volumes of companion texts such as Holland's The Moth Book or Kimball's Lepidoptera of Florida, now we have access to another extremely useful and updated identification guide for moths. An added advantage is that many species not illustrated in these other references are included here; a total in excess of 1300 species is treated.

One must quickly realize the impossibility of the guide to give discussions and illustrations for ALL of the species present in the region covered. Most of the more common and larger species are presented. While some of us might have enjoyed seeing more of the microheterocera, we must also realize the primary intention of the author to introduce new faces to the wonders of lepidoptera. The text accomplishes this quite well; I would also hope that the book will stimulate more individuals to undertake the study of moths. If anything, butterflies have and will always be at the forefront of the study of lepidoptera, primarily because of their high visibility and the relative ease of identification. The sheer numbers one first faces when beginning a collection of moths is intimidating, and I suppose that some of these individuals quickly lose heart because of the difficulty in trying to identify specimens.

Most of the individual states covered in the guide harbor less than 150 different butterfly/skipper species; most of these same states will similarly provide residency for OVER 2000 different species of moths!

As a result, butterflies tend to be over-studied and moths tend to be greatly ignored by many amateur collectors. Hopefully, this text will provide a renewed spark of interest and enthusiasm for the "old hands" as well as to encourage the interest of those just beginning to learn about lepidoptera.

Excellent discussion of collecting techniques is given - perhaps the best synopsis ever presented in a field guide. The usual attention to areas such as anatomy and life cycle is given in good detail, supplemented by an adequate glossary covering terminology. I was most pleased to find a section on rearing specimens - an area of moth study where ANY individual can help make contributions to the knowledge of lepidoptera, as many life histories are at best poorly known. A great deal of thought went into the section on collecting and preserving specimens; this is presented in informative yet concise fashion by Dr. Covell. I am equally certain that the author spent considerable time and effort selecting the species used for discussion and illustration.

The nomenclature used is up-to-date and generally follows the recent Hodges, et al, MONA checklist (1983). A glaring exception is the use of the genus Grammia for the popular tiger moth genus most of us know as Apantesis. However, I feel confident that his use of this hinged on some debate of what was going to be employed in the MONA checklist, and that at the time the guide was being prepared, this issue was not resolved. While the nomenclature basically follows the MONA list, neither the text nor the plates follow this arrangement, although the present classification scheme is presented in the introduction section. It should be obvious to any reader that the progression employed is that of simple convenience from large to small species, consistent with the intention of the guide. My ONLY real complaint is that several of the B & W plates are nearly illegible, but this is clearly the fault of the publisher.

In general, the text is relatively error free when considering the amount of material presented and the region covered by the author. I am aware of a few errors: the

figures on plate 12 for Cisthene tenuifascia and C. kentuckiensis are reversed; Egira "alternata" should be E. alternans on both plate 22 (fig. 11) and in the text on page 107; lastly, figure 8 on plate 34 is given as Catocala miranda when it should be Catocala orba. (This reviewer SENT the author the specimen figured, thinking it WAS C. miranda. The error here obviously was MINE, NOT the author's.) However, the diagnostic key for separation of the two species on page 313 is reversed for the two species; it is C. miranda which should have the dark median dash on the inner margin, NOT C. orba; C. miranda typically is a smaller species than orba. Figure 12, plate 12 for Pygarctia abdominalis states "reared", yet the text indicated the larval host plant as "unrecorded". This serves as a classic example of the importance of recording the host plant when labelling a reared specimen. There are many similar examples in both museum and private collections whereby a collector has taken time to note that the specimen was reared, but has totally ignored the importance of ON WHAT ?? This is a considerable source of irritation and frustration to subsequent researchers, especially in cases where the ranges are projected beyond current known range (using Florida as an example, an area where I am relatively familiar with the respective species) or inadvertently omitted, but these are generally understandable exceptions.

Concluding, I find the guide to be one of the most useful in recent years, and any serious lepidopterist should welcome this addition to their library. No text is perfect, but this comes pretty close considering the amount of material covered and the area represented. While it might not rate an "A+", it surely rates an "A". My hard-bound copy will stay at home, but the paperback will follow me often into the field here in Florida on my "expeditions"! Dave Baggett

[Ed. Note] Nor can we overlook the first acknowledgment of the Southern Lepidopterists' Society in a text...thanks for remembering us, Charlie!

## OBITUARY

MR. HAROLD LAVERNE KING passed away on January 27, 1985 at his home in Sarasota, Fla. following an extended illness. A charter member of The Lepidopterists' Society and a Research Associate of the Florida State Collection of Arthropods since 1969, "Verne" collected Lepidoptera as an avocation much of his lifetime with a special interest in Lycaenidae. He made a significant contribution to our knowledge of the Lepidoptera of Florida, other parts of the United States and especially of Mexico and Central America. His large, neatly prepared, accurately identified collection, which included some very rarely collected Neotropical species, had been deposited in the FSCA prior to his death at the age of 85. Mr. King was a commercial artist and for many years owned and operated King of Sarasota Advertising Service, which involved a billboard business throughout much of Florida and southern Georgia. He is survived by his wife, Suejette, and three children by an earlier marriage: Mrs. Carolyn Clough, Jr., Ms. Polly Ann King, and Mr. William C. King.

Howard V. Weems, Jr.  
Taxonomic Entomologist & Curator  
Florida State Collection of Arthropods

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## RESEARCH REQUESTS

Records are needed from anyone with data on species collected in Florida to help improve knowledge of distributional records of Floridian species in conjunction with revision plans for Kimball's Lepidoptera of Florida. contact: Dave Baggett, 14406 N. 22nd. St. #169, Lutz, Florida 33549.

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Dr. Stuart J. Ramos, Dean of Students, Universidad de Puerto Rico, College Station, Mayaguez, PR 00708 is planning an extensive study of the Pierid genus Eurena, and would appreciate information from our members. He also badly needs immature material for the study, and would be extremely pleased to have help with this from the mainland. If you have access to preserved material, or the opportunity to obtain livestock during 1985, please contact him.

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## ZONE REPORTS:

ZONE I: TEXAS, COORDINATORS: Ed Knudson, 808 Woodstock, Bellaire 77401; Mike Rickard, 6550 Hillcroft #201, Houston 77081. Data on moths should go to Ed and data on butterflies to Mike.

Tom Kral took the third Texas specimen of Letis xylia (a perfect male) in a bait trap Oct. 21, 1984 at the Santa Ana Refuge, Hidalgo County. This moth looks much like a small black witch moth.

Ed Knudson reported the following: Ft. Bend Co., Brazos Bend St. Pk. 26 Feb.: Phaneta ambodaidalea, Ancyliis muricana (Tortricidae); Ceratonyx satanaria (Geometridae); Elaphria versicolor, E. exesa, Orthosia alurina (Noctuidae). Huntsville State Park, Walker Co., 11 March: Cameraria ulmella (Gracillariidae); Eupithecia peckorum (Geometridae); Zale galbanata, Z. minerea, Z. phaeocapna, Z. confusa, Z. bethunei, Z. calycanthata, Pyreferra pettiti, Cerastis tenebrifera (Noctuidae). Lake Tawakoni, Hunt Co., 12 March: Caloptilia atomosella, Cameraria affinis (Gracillariidae); Eutolype electilis, Orthosia hibisci (Noctuidae). Palmetto State Park, Gonzalez Co., 23 March: Phyllonorycter basistrigella (Gracillariidae); Ethmia delliella, E. bittenella (Oecophoridae); Epermenia sp. (Epermeniidae); Usingeriessa brunnilalis (Pyrilidae); Metanema inatomaria (Geometridae); Eutelia pulcherrima, Acronicta americana, A. tota, A. connecta, A. atristrigata, A. retardata, Harrisimemna trisignata (Noctuidae). Garner State Park, Uvalde Co., 25 March. Bucculatrix staintonella (Lyonetiidae); Ellabella editha (Copromorphidae); Oidaematophorus longifrons (Pterophoridae); Lomographa glomeraria, Exilis ophiurus, Drepanulatrix n.sp., Sicya morsicaria, Synaxis triangulata, Anticlea multiferata (Geometridae); Furcula borealis, Heterocampa belfragei (Notodontidae); Toxonprucha psegmapteryx, Zale edusina, Z. colorado, Tripudia inquaesita, Stiriodes perflava, Protorthodes orobia (Noctuidae).

ZONE II: ALA., LA., MISS., TENN. COORDINATORS: Vernon Brou, 137 Jack Loyd Rd., Abaita Springs, La. 70420; Bryant Mather, 213 Mt. Salus Dr., Clinton, Miss. 39056; Charles Watson, 1339 Watauga St., Kingsport, Tn. 37660; John Hyatt, 439 Forest Hills Dr., Kingsport, Tn. 37663.

LOUISIANA: Brou reported the following, all as NEW STATE RECORDS: Eurythmia hospitella, Diaphania indica, Paraponyx diminutalis, Tetralopha scortealis, T. militella, T. humerella, Homoeosoma electellum, Vitula edmandsii, Achroia grisella, Scirpophaga repugnatalis, Peoria longipalpella, P. bipartitella, P. roseotinctella, Tampa dimediatella, Homosassa platella, Atascosa glareosella (Pyrilidae); Parastichtis discivaria, Minofala instans (Noctuidae); Gonioterma mistrella (Oecophoridae); 5 new undescribed Dichomeris sp., Dichomeris vaciniella, Deltophora sella, Helcystogramma melanocarpa, Isophrictis rudbeckiella, Chionodes pereyra (Gelech.).

ZONE IV: FLORIDA, COORDINATOR Dave Baggett, 14406 N. 22nd Street #169, Lutz, Fl 33549

Dr. Larry Brown took a small series of Synanthedon dominicki at the USF ecological area in Tampa. This rare sesiid was only known from the male holotype from the Wedge Plantation, Charleston Co., S.C.! Don Lafontaine found a specimen of Euxoa violaris in the USNM which is a Florida state record. The specimen was from Ocean City. Eric Metzler found Cycnia pudens at Torreya St. Pk. for another new Florida record.

The following are new county records for Catocala species. C. sappho, Citrus Co., by Rick Gillmore; C. jair, Duval Co., by Jeff Slotten; C. gracilis, Citrus Co., by Dave Baggett; C. connubialis, Duval Co., by Slotten. Thirty five different Catocala species have been taken to date in excellent spring collecting. Bait has been very productive probably due to dry conditions.

Baggett reported that Pontia protodice is having a population explosion locally with hundreds along the roadsides- everywhere. Mike McInnis took Megathymus cofaqui, Yankeetown, Levy County. In early February Woody Dow and Dennis Profant reported a good flight of Hemileuca maia near Deltona, this is unusually late for this species even in Florida.

[I have had a very slow trickle of info come in. I hope to have #2 out in SIX weeks, so send in all that information you are setting on. Sorry this is so late! -Ron ]

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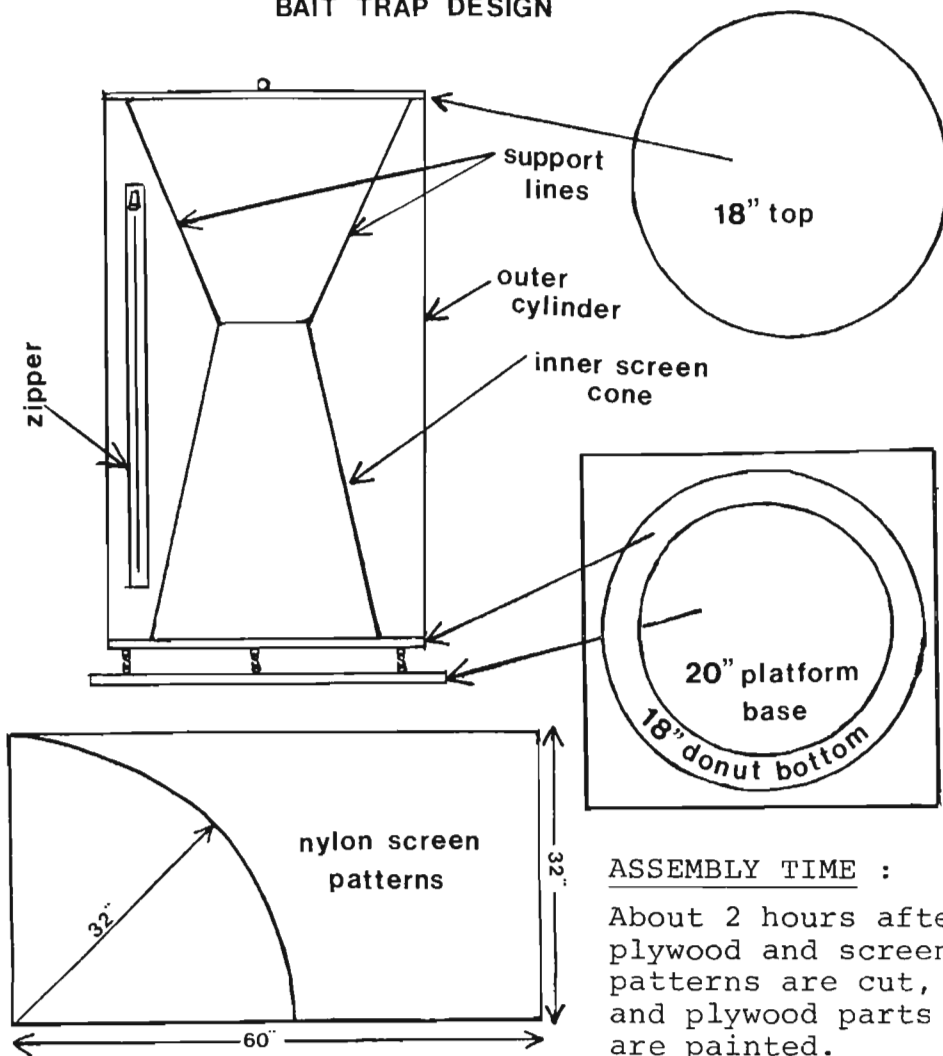
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THE SOUTHERN LEPIDOPTERISTS' NEWS  
C/O the Editor, Ron Gatrell  
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GOOSE CREEK, S.C. 29445

Dr. Lee D. Miller  
Allyn Museum  
3701 Bay Shore Road  
Sarasota, FL 33580



## BAIT TRAP DESIGN



## MATERIALS NEEDED :

H.D. Staple Gun  
Large sewing needle  
10# mono fishing line  
wire coat hanger  
one 22" nylon zipper  
four small 's' hooks  
8 small screw eyes  
one large screw eye  
20' 3/16" nylon rope  
8' x 32" nylon screen  
2' x 6' x 5/8" plywood  
jig saw or coping saw  
screwdriver, scissors  
spray enamel paint  
safety pins, pliers  
1" deep plastic dish

## BAIT RECIPE :

3 cans beer (not Lite)  
2 lbs. white sugar  
½ jar unsulfured molasses  
3 cups of crushed fruit  
(apples, bananas, peaches  
mixed to pulp in blender)  
Mix above together and  
place in a lightly-capped  
gallon milk jug to fer-  
ment for several days be-  
fore use. Add about ½"  
to bait dish, placed on  
platform below cone.

## ASSEMBLY TIME :

About 2 hours after  
plywood and screen  
patterns are cut,  
and plywood parts  
are painted.

**ASSEMBLY INSTRUCTIONS :** First, mark and cut the screen and plywood to the indicated specifications, noting that the "donut" plywood bottom and the ¼-circle screen cone patterns have simply been superimposed to conserve space in the illustrations above. You should have three plywood pieces - a 20" square, an 18" dia. circle, and an 18" dia. donut with 15½" opening - and two pieces of screen - a 32" x 60" rectangle used to fashion the outer screen cylinder of the trap, and a ¼-circle piece of 32" radius which is used to make the entrance cone of the trap. The plywood parts should be spray-painted for protection from the rain. Using some safety pins, roll the screen cone together and pin along the overlapping straight edges to hold shape. Form a hoop of 5" diameter from the coat hanger wire (or use a 5" macrame hoop); slide down the top of the cone, trimming off apparent excess while leaving about ½" overlap for attaching the hoop to the top of the screen cone by sewing. Use the mono fishing line for thread - it is strong and weatherproof. After attaching the hoop, sew the seam shut along the side. Slide the cone through the donut interior, then staple the screen to the donut along the inside edge. Trim to leave about 1" excess, and staple around the bottom of the donut. Then begin stapling the outer screen cylinder portion around the outer edge of the plywood donut. Finished with this, staple three 18" pieces of fishing line equidistant around the inner perimeter of the plywood top, attaching at one end only. These pieces of line form the stays to support the cone from the hoop. Proceed to

staple the opposite end of the screen cylinder to the plywood top, tacking the screen in place around the top. At this point, the basic shape of the trap should be apparent. Attach the large screw eye to the center of the top as shown, and suspend the trap with some line so that it hangs about waist-high. By suspending the trap at this point, you can insure neat seams on the outside as you sew it shut, after fastening the stays to the hoop for support inside. (I prefer to use snaps or safety pins at the end of the stays, which allow you to unfasten the inner cone after assembly, which facilitates cleaning of the trap.) After the cylinder seam is sewn shut (there should be several inches of sufficient overlap, I position the zipper in place over the center of the sewn seam, stretch taut, and fasten in position with the safety pins at each end. Using scissors, cut an opening through the screen seam about  $\frac{1}{4}$ " wide and 22" long to facilitate the zipper. A nylon zipper is used because a metal one will corrode when used outdoors. Sew the zipper in place; the overlapped screen seam reinforces the area where the zipper is attached. The screen portion of the trap should now be completed. To attach the bait dish platform (the 20" square), first attach the small screw eyes 90° ( $\frac{1}{4}$  circle) apart around the bottom of the wooden donut (see diagram); insert one end of an 's' hook in each screw eye and bend shut around eyes with pliers. Sizing the precise positioning of the screw eye attachment to the donut, mark these positions accordingly on the platform base, and insert the remaining screw eyes in the base. The wooden bait platform will hang easily suspended by attaching the free ends of the 's' hooks hanging from the donut to the screw eyes positioned on the platform. After attaching the nylon rope to the large screw eye at the top of the trap, the trap is now ready for use. The rope is used to suspend the trap from a convenient tree branch, etc., outdoors, and also is used to tie up the trap for transportation purposes, as the trap parts will collapse neatly for this purpose. A dish of fermented fruit bait is placed on the platform directly below the cone. I should caution you that bait should always be stored LOOSELY capped, to prevent fermentation gases from blowing up the container. If this happens while transporting bait in the car, you'll never forget the ensuing mess! These traps are also ideal for use with pheromone baits, and by inverting the trap with the platform removed, a 15-watt blacklight can be hung in such a fashion that it can be suspended half-way down the cone to form a portable light trap. The applications for insect-collecting are very much underestimated by most collectors. Live traps are time-savers, requiring minimal daily attendance, and also allow for selective collecting; take what you need and release the rest. The bait platform can be easily removed, and the screen cone can be dropped out the bottom after undoing the support lines above; the trap can be shaken vigorously and the bugs not wanted will fall out the bottom. You need to be a bit careful, though, as traps will also collect wasps and hornets. These normally will be able to leave the trap on their own (a lot smarter than leps, apparently) at dusk, but once in a while a "dummy" will get lost. I find that you can easily jar wanted specimens through the zipper access even with wasps present, since they normally crawl or fly around, undisturbed by the affair, but if you accidentally press one between your arm and the screen mesh while reaching for a specimen, expect the consequences; i.e., look first! Bait traps are extremely useful for collecting certain Sphingidae, most Noctuidae (very effective for the popular Catocala moths), most Nymphalid butterflies, most Satyrid butterflies, and also occasionally collect Hesperidae and Lycaenidae. They are equally effective for additional insect orders. You may do well in one area and poorly in another, so vary locations.